July 13, 2018

Dear Dr. Phillip Steindel and Referees,

We thank the editor and the three reviewers again for your constructive and very helpful comments. Accordingly, we made corrections to the text and the figure legends corresponding to the suggestions.

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**Editorial comments:**

Note that some formatting changes have already been made.  
1. Per your response to Reviewer 1, could you give more specific details as to how old mice should be for this protocol to be successful?

In our experience, we can obtain healthy spinal cord slices successfully from neonatal as well as 3-10 weeks old SD rats. Thus, for this protocol to be successful, we recommend using SD rats less than 10 weeks old.

2. 3.3.5 (current numbering): Can you indicate how the cord is attached to the agar block (not just how it fits into it)? Superglue?

We apologize for not making this statement clear. We have now added the following description in the text in section 3.3.5: ‘use superglue to adhere the spinal cord tissue to the block’.

3. The legend for Figure 3B+C no longer applies to the current figure. Additionally; why did you remove the original 3B+C?

We thank the editor for pointing this out. We have now updated the legend for Figure 3B+C in the revised version as follows ‘**B.** Representative traces of sEPSCsrecorded from SG neurons at -70 mV in the absence and presence of 50 μM APV and 20 μM CNQX**.** Lower consecutive traces, which are shown in an expanded time scale before (left) and under (right) the action of APV and CNQX, correspond to a period indicated by a bar shown below the chart recording. **C.** Representative traces of sIPSCsrecorded from SG neurons in the absence and presence of 10 μM bicuculline and 1 μM strychnine at 0 mV.’

As Reviewer 1 suggested us to add traces showing EPSCs and IPSCs could be confirmed with specific blockers, we substituted the original 3B+C with traces showing that EPSCs and IPSCs could be confirmed with APV and CNQX, bicuculline and strychnine, respectively.